

# Imaging solutions: Driving agri-food tech towards sustainability

Today's global food production system faces significant challenges crucial to sustaining our planet's fast-growing population. This growth requires a significant increase in the food supply, with estimates suggesting a need to boost food production by up to 70% to meet rising demand. At the same time, the need for sustainable farming practices has never been more critical. The agriculture sector contributes approximately 24% of global greenhouse gas emissions and plays a significant role in water usage and habitat loss.

These environmental impacts conflict with the European Union's sustainability goals, which aim for a climate-neutral continent by 2050. Climate change intensifies the challenges faced by the agri-food sector, heightening crops' vulnerability to pests, diseases, and extreme weather events. Additionally, roughly one-third of all food produced for human consumption is lost or wasted. To address these challenges, balancing increased food production with environmental sustainability goals is essential for the future of global food systems, highlighting the need for innovative approaches and solutions.

## Photonics at the heart of the solution

Photonics and imaging technology, specifically, are crucial to addressing these challenges, offering innovative tools that transform the agri-food tech sector. Photonics-based instruments emit light toward a target, interacting with the material in ways specific to its composition and structure. The light may be absorbed, reflected, transmitted, or cause the material to fluoresce. Advanced sensors capture these interactions, providing detailed information about the target's chemical, physical, and biological makeup through sophisticated analysis.



The main benefits of photonic solutions are their non-contact, fast, and primarily non-destructive testing nature. This non-invasive approach allows for real-time monitoring and inspection of agricultural products and foodstuffs, enabling the detection of diseases, nutritional content, moisture levels, and other critical parameters without compromising sample integrity, as shown in Table 1 below.

**Table 1. Overview of non-destructive methods for evaluating fruit ripeness and their correlation with internal characteristics.** <sup>[1]</sup>

Abbreviations: SSC (Soluble Solids Content), DM (Dry Matter), MC (Moisture Content), TTA (Titratable Acidity), TSS (Total Soluble Solids), and WC (Water Content). Each numbers in the table are referenced in the reference section at the end of the article.

	Colorimetry	Visible imaging	Spectroscopy	Fluorescence	Hyper spectral imaging	Multispectral imaging
Apple	Color [21]	Color [22]	Chlorophyll [23], Anthocyanins [24], Carotenoids [24], Flavonols [25], SSC [26], Firmness [27]	Chlorophyll [28], Anthocyanins [29], Flavonols [29], Firmness [29], SSC [29]	Firmness [30], SSC [31]	Firmness [32], SSC [33]
Pear			Firmness [34], SSC [35]		SSC [36]	
Peach	Color [37]		Firmness [38], Chlorophyll [39], Color [37]	Firmness [40]	Firmness [41]	Firmness [9], SSC
Avocado			MC [42], DM [42]		DM [43]	
Nectarine	Color [44]		SSC [45], Firmness [45]	Firmness [40]		
Mango	Color [46]		DM [47], Starch [48], SSC [47], Color [49], Firmness [50]		Firmness [8], SSC [8], WC [8]	
Banana	Color [51]	Color [51]	TSS [52], Chlorophyll [53]		Firmness [54], TSS [54]	
Tomato	Color [55], Firmness [56], TSS [57]	Color [58], Firmness [59]	Lycopene [60], SSC [61]	Chlorophyll [62]		Phenolic [63], Lycopene [63]
Melon			SSC [64]			
Mandarin			TTA [65], SSC[66], Firmness [67], DM [68]			
Cherry	Color [69]		Firmness [70], SSC [71]			
Strawberry			Color [72], TTS [73], Firmness [72], TTA [73]		Firmness [74], TSS [75], TTA [75]	SSC [76], Firmness [77]
Apricot			SSC [78], Firmness [78], TTA [78]			
Kiwifruit			TSS [79], SSC [79], Firmness [80], DM [81], Starch content [79]			
Persimmon			SSC [82]		Firmness [83]	
Grape			SSC [84], TTA [84], Anthocyanin [85]	Chlorophyll [86], Anthocyanin [86], TSS [86], Flavonols [87]	SSC [88], TTA [88]	
Pineapple		Color [89]	DM [90], SSC [91]			
Plum			Firmness [92], SSC [93], Color [92]			



By leveraging light-matter interactions, photonics enables precise, efficient, and sustainable management practices in the agri-food sector, promoting higher yields, reduced waste, and enhanced food safety.

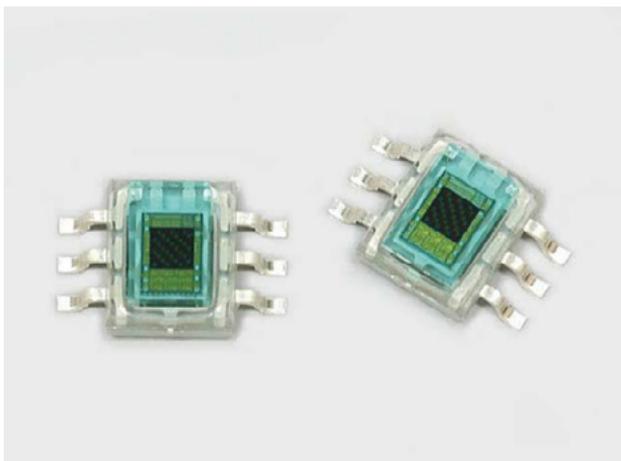
The following non-destructive techniques are instrumental in advancing the agri-food tech sector, providing precise and efficient tools for assessing and improving the quality, safety, and sustainability of agricultural products and food:

1. Colorimetry & visible imaging
2. Visible & near-infrared (VNIR) spectroscopy
3. Fluorescence imaging
4. Hyperspectral imaging
5. Multispectral imaging

## 1. Colorimetry & visible imaging

Colorimetry and visible imaging are essential techniques in the agricultural and food technology (agri-food tech) sector. They provide a straightforward, non-destructive means of assessing the quality and characteristics of food and farming products by measuring the intensity of light reflected from an object in the visible spectrum (approximately 400-700 nm), offering insights based on color variations and patterns.

Sensor technological advancements have enabled a shift from human evaluation to measurements in the CIELAB color space ( $L^*$ ,  $a^*$ ,  $b^*$ ), which offers highly accurate and consistent color measurements. The rise of RGB sensors, capable of simultaneously measuring RGB colors on a 2-D area with digital output, has further boosted colorimetry and visible imaging by offering a straightforward and cost-effective setup for many applications (see, for example, [Hamamatsu S9706](#)<sup>[2]</sup>)



Hamamatsu's S9706.

In agri-food tech, colorimetry and visible imaging are extensively used for quality control and sorting processes. For example, in the fruit and vegetable industry, these technologies enable the automated sorting of products based on ripeness, color uniformity, and surface defects. In apple orchards, colorimetry determines the optimal harvest time by assessing the apple color development, ensuring that only fruits meeting specific maturity criteria are picked. This maximizes the quality and taste of the harvested fruits and reduces waste by minimizing the picking of underripe or overripe produce. Similarly, in grain quality assessment, visible imaging helps to identify and segregate grains infected by fungi or showing signs of sprouting, thereby improving the safety and quality of the grain supply.

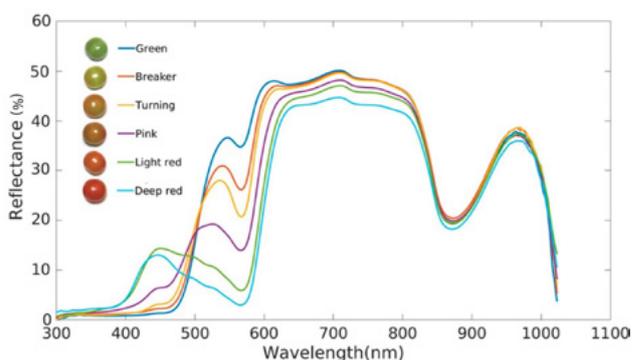
## 2. Visible & near-infrared spectroscopy

Compared to the previous technique, visible and near-infrared (VNIR) spectroscopy offers a more nuanced and in-depth analysis tool in the agri-food tech sector. VNIR spectroscopy extends analysis into the near-infrared range (approximately 400-2500 nm), tapping into the unique ways materials interact with light across both visible and NIR wavelengths. This spectral range is particularly insightful because molecular vibrations and overtones, indicative of chemical bonds in organic compounds, strongly absorb NIR light, allowing precise identification of chemical compositions and concentrations within agricultural products and foods.

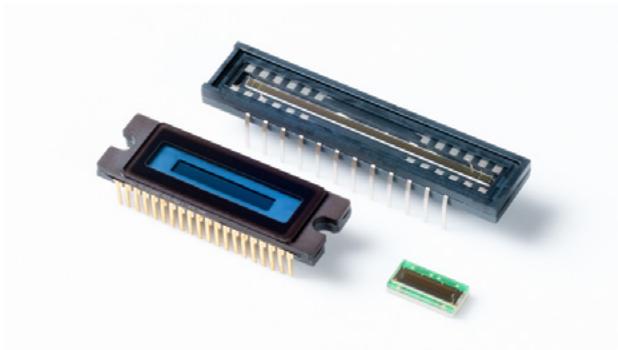
VNIR spectroscopy enables the detection and quantification of organic compounds and moisture content with high sensitivity and specificity. For example, in precision agriculture, VNIR spectroscopy can analyze crop foliage to determine nitrogen levels, directly impacting fertilization decisions crucial for crop yield optimization.

As seen in Graph 1 below, in food quality assessment, VNIR spectroscopy is used to quantify the sweetness of fruits such as apples and berries by measuring sugar content directly through their skin. This allows for sorting into different quality grades without damaging the produce.

**Graph 1: Typical progressive change of reflectance spectra at different ripening stages of tomato**<sup>[3]</sup>



The same physical principle can be used to study the in-line defectivity of food products, combining both sensors working in the visible spectrum ([Hamamatsu CCD Sensors](#)<sup>[4]</sup>) and sensors working in the infrared spectrum ([Hamamatsu InGaAs Sensors](#)<sup>[5]</sup>).

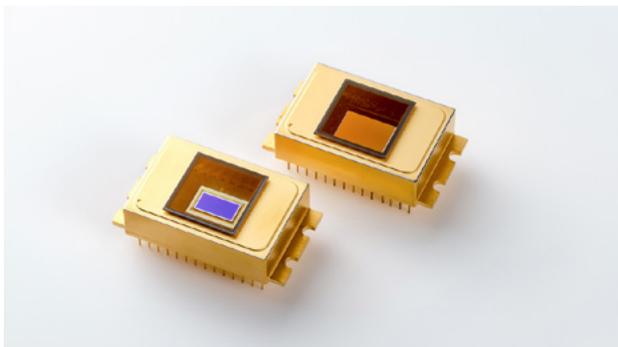


Hamamatsu's CCD sensors

Thus, VNIR spectroscopy's ability to provide rapid, non-destructive analysis across various samples makes it an invaluable tool for real-time monitoring and quality control in food production.

### 3. Fluorescence imaging

Building on the depth of analysis provided by VNIR spectroscopy, fluorescence imaging introduces a dynamic dimension to the agri-food tech sector's toolkit. Fluorescence imaging relies on the ability of certain substances to absorb light at one wavelength and emit it at another, longer wavelength. To detect even small amounts of markers, accurate and highly sensitive sensors at different wavelengths are mandatory, like [Hamamatsu InGaAs area sensors](#) <sup>[6]</sup>.

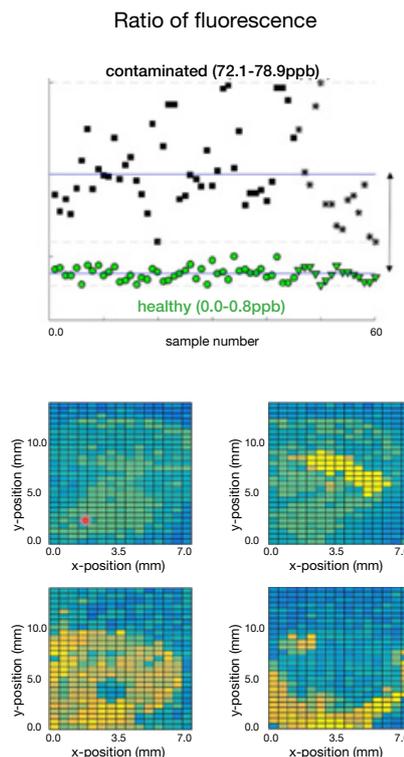


Hamamatsu's InGaAs area sensors

Fluorescence imaging applications in agri-food tech range from detecting microbial contamination in food products to assessing plant health. For instance, specific pathogens on fruit skins can be identified through their distinct fluorescence signatures, enabling early detection of spoilage and contamination without physical contact. Similarly, fluorescence imaging can identify areas of stress in crops caused by drought or disease by observing changes in chlorophyll fluorescence, which correlates with the plant's photosynthetic activity and overall health.

A particularly impactful application has been the rapid screening of mycotoxins in grains and nuts, posing significant health risks. Fluorescence imaging can detect these toxins with high sensitivity, often enabling detection levels down to parts per billion. This capability significantly enhances food safety protocols, allowing for the precise identification and removal of contaminated products from the supply chain.

**Graph 2: Fluorescence spectroscopy enabling toxin detection**<sup>[7]</sup>



### 4. Hyperspectral imaging

With modern sensors, it is now possible to combine the aforementioned techniques and collect a complete continuous spectrum for each pixel, using it for hyperspectral imaging. This comprehensive spectral data enables the detailed characterization of agricultural products and foods' chemical and physical makeup by detecting subtle differences in light absorption and reflection patterns. It can identify and quantify various substances and conditions, from crops' moisture content and protein levels to subtle signs of diseases and nutrient deficiencies not visible to the naked eye.

In precision agriculture, hyperspectral imaging mounted on unmanned aerial vehicles (UAVs) can map variability in soil properties across a field, enabling targeted fertil-

ization and irrigation that enhance crop yield and reduce resource use. In food quality assessment, this technology can detect foreign bodies or contamination, such as bits of plastic or metal, and monitor the freshness and ripeness of produce without damaging it.

### Graph 3: Hyperspectral imaging assessment of different coffee bean types.

Hamamatsu's Energetiq tuneable light sources (LDTLS<sup>®</sup>) and InGaAs cameras were utilized to demonstrate the imaging.



Due to the level of detail provided by hyperspectral imaging and the availability of integrated hyperspectral cameras, such as the SXGA InGaAs Camera <sup>[8]</sup>, its application is continually expanding. It offers a potent tool for advancing the efficiency, sustainability, and safety of food production.

Multispectral imaging's balance between spectral detail and operational simplicity makes it an invaluable tool in agri-food technology. Its focused, rapid analysis capabilities facilitate sustainable farming practices and enhance food quality.



Hamamatsu's SXGA InGaAs Camera

## Imaging tools are critical to a more sustainable agri-food tech sector

As global food demands rise and the need for sustainable agriculture intensifies, imaging technologies emerge as essential allies in the agri-food tech sector. These technologies serve a dual purpose: enhancing productivity and sustainability in food production and processing. They enable non-destructive, precise analysis of crops and food products, offering valuable insights into quality, safety, and environmental impact. They facilitate crop health monitoring, resource optimization, and waste reduction and address critical challenges in the agri-food sector. Integrating imaging technologies is key to achieving higher yields, reducing environmental footprint, and ensuring food safety and quality.

As these technologies evolve and become more accessible, they will play an increasingly significant role in sustainably meeting the world's food production needs, addressing immediate agricultural challenges, and paving the way for future advancements in global food supply and environmental care.

Please contact Hamamatsu Photonics Europe for advice on your agri-food needs: [info@hamamatsu.eu](mailto:info@hamamatsu.eu)

## References

<sup>11</sup>AB. Li, J. Lecourt, and G. Bishop, "Advances in Non-Destructive Early Assessment of Fruit Ripeness towards Defining Optimal Time of Harvest and Yield Prediction—A Review," *Plants*, vol. 7, no. 1, p. 3, Jan. 2018, doi: <https://www.mdpi.com/2223-7747/7/1/3#B101-plants-07-00003>

[8] Sivakumar, S.S.; Qiao, J.; Wang, N.; Gariépy, Y.; Raghavan, G.S.V.; McGill, J. Detecting maturity parameters of mango using hyperspectral imaging technique. In Proceedings of the 2006 ASAE Annual Meeting, Portland, OR, USA, 9–12 July 2006. [Google Scholar]

[9] Lleó, L.; Barreiro, P.; Ruiz-Altisent, M.; Herrero, A. Multispectral images of peach related to firmness and maturity at harvest. *J. Food Eng.* 2009, 93, 229–235. [Google Scholar] [CrossRef] [Green Version]

[21] Zude, M. Comparison of indices and multivariate models to non-destructively predict the fruit chlorophyll by means of visible spectrometry in apple fruit. *Anal. Chim. Acta* 2003, 481, 119–126. [Google Scholar] [CrossRef]

[22] Merzlyak, M.N.; Solovchenko, A.E.; Gitelson, A.A. Reflectance spectral features and non-destructive estimation of chlorophyll, carotenoid and anthocyanin content in apple fruit. *Postharvest Biol. Technol.* 2003, 27, 197–211. [Google Scholar] [CrossRef]

[23] Solovchenko, A.E.; Chivkunova, O.B.; Gitelson, A.A.; Merzlyak, M.N. Non-destructive estimation pigment content, ripening, quality and damage in apple fruit with spectral reflectance in the visible range. *Fresh Prod.* 2010, 4, 91–102. [Google Scholar]

[24] Peirs, A.; Scheerlinck, N.; Nicolai, B.M. Temperature compensation for near infrared reflectance measurement of apple fruit soluble solids contents. *Postharvest Biol. Technol.* 2003, 30, 233–248. [Google Scholar] [CrossRef]

[25] Zude, M.; Herold, B.; Roger, J.M.; Bellon-Maurel, V.; Landahl, S. Non-destructive tests on the prediction of apple fruit flesh firmness and soluble solids content on tree and in shelf life. *J. Food Eng.* 2006, 77, 254–260. [Google Scholar] [CrossRef]

[26] Zude-Sasse, M.; Truppel, I.; Herold, B. An approach to non-destructive apple fruit chlorophyll determination. *Postharvest Biol. Technol.* 2002, 25, 123–133. [Google Scholar] [CrossRef]

[27] Betemps, D.L.; Fachinello, J.C.; Galarça, S.P.; Portela, N.M.; Remorini, D.; Massai, R.; Agati, G. Non-destructive evaluation of ripening and quality traits in apples using a multiparametric fluorescence sensor. *J. Sci. Food Agric.* 2012, 92, 1855–1864. [Google Scholar] [CrossRef] [PubMed]

[28] Peng, Y.; Lu, R. Analysis of spatially resolved hyperspectral scattering images for assessing apple fruit firmness and soluble solids content. *Postharvest Biol. Technol.* 2008, 48, 52–62. [Google Scholar] [CrossRef]

[29] Mendoza, F.; Lu, R.; Ariana, D.; Cen, H.; Bailey, B. Integrated spectral and image analysis of hyperspectral scattering data for prediction of apple fruit firmness and soluble solids content. *Postharvest Biol. Technol.* 2011, 62, 149–160. [Google Scholar] [CrossRef]

[30] Peng, Y.; Lu, R. An LCTF-based multispectral imaging system for estimation of apple fruit firmness: Part II. Selection of optimal wavelengths and development of prediction models. *Trans. ASABE* 2006, 49, 269–275. [Google Scholar] [CrossRef]

[31] Peng, Y.; Lu, R. Prediction of apple fruit firmness and soluble solids content using characteristics of multispectral scattering images. *J. Food Eng.* 2007, 82, 142–152. [Google Scholar] [CrossRef]

[32] Cavaco, A.M.; Pinto, P.; Antunes, M.D.; Silva, J.M.; da Guerra, R. "Rocha" pear firmness predicted by a Vis/NIR segmented model. *Postharvest Biol. Technol.* 2009, 51, 311–319. [Google Scholar] [CrossRef]

[33] Shao, Y.; Bao, Y.; He, Y. Visible/Near-Infrared spectra for linear and nonlinear calibrations: A case to predict soluble solids contents and pH value in peach. *Food Bioprocess Technol.* 2011, 4, 1376–1383. [Google Scholar] [CrossRef]

[34] Tiansheng, H.; Jun, Q.; Wang, N.; Ngadi, M.O.; Zuoxi, Z.; Zhen, L. Non-destructive inspection of Chinese pear quality based on hyperspectral imaging technique. *Trans. Chin. Soc. Agric. Eng.* 2007, 2, 29. [Google Scholar]

[35] Ferrer, A.; Remon, S.; Negueruela, A.I.; Oria, R. Changes during the ripening of the very late season Spanish peach cultivar Calanda: Feasibility of using CIELAB coordinates as maturity indices. *Sci. Hortic.* 2005, 105, 435–446. [Google Scholar] [CrossRef]

[36] Lafuente, V.; Herrera, L.J.; Pérez, M.D.M.; Val, J.; Negueruela, I. Firmness prediction in *Prunus persica* 'Calrico' peaches by visible/short-wave near infrared spectroscopy and acoustic measurements using optimised linear and non-linear chemometric models. *J. Sci. Food Agric.* 2015, 95, 2033–2040. [Google Scholar] [CrossRef] [PubMed]

[37] Ziosi, V.; Noferini, M.; Fiori, G.; Tadiello, A.; Trainotti, L.; Casadoro, G.; Costa, G. A new index based on vis spectroscopy to characterize the progression of ripening in peach fruit. *Postharvest Biol. Technol.* 2008, 49, 319–329. [Google Scholar] [CrossRef]

[38] Bodría, L.; Fiala, M.; Guidetti, R.; Oberti, R. Optical techniques to estimate the ripeness of red-pigmented fruits. *Trans. ASAE* 2004, 47, 815–820. [Google Scholar] [CrossRef]

[39] Lu, R.; Peng, Y. Hyperspectral Scattering for assessing Peach Fruit Firmness. *Biosyst. Eng.* 2006, 93, 161–171. [Google Scholar] [CrossRef]

[40] Olarewaju, O.O.; Bertling, I.; Magwaza, L.S. Non-destructive evaluation of avocado fruit maturity using near infrared spectroscopy and PLS regression models. *Sci. Hortic.* 2016, 199, 229–236. [Google Scholar] [CrossRef]

[41] Girod, D.; Landry, J.A.; Doyon, G.; Osuna-García, J.A.; Salazar-García, S.; Goenaga-Portela, R. Evaluating Hass avocado maturity using hyperspectral imaging. *Caribb. Food Crops Soc. Proc.* 2008, 44, 144–154. [Google Scholar]

[42] Luchsinger, L.E.; Walsh, C.S. Development of an objective and non-destructive harvest maturity index for peaches and nectarines. *Acta Hortic.* 1998, 465, 679–688. [Google Scholar] [CrossRef]

[43] Pérez-Marín, D.; Sánchez, M.T.; Paz, P.; Soriano, M.A.; Guerrero, J.E.; Garrido-Varo, A. Non-destructive determination of quality parameters in nectarines during on-tree ripening and postharvest storage. *Postharvest Biol. Technol.* 2009, 52, 180–188. [Google Scholar] [CrossRef]

[44] Jha, S.N.; Chopra, S.; Kingsly, A.R.P. Modeling of color values for nondestructive evaluation of maturity of mango. *J. Food Eng.* 2007, 78, 22–26. [Google Scholar] [CrossRef]

[45] Subedi, P.; Walsh, K.; Purdy, P. Determination of optimum maturity stages of mangoes using fruit spectral signatures. *Int. Soc. Hortic. Sci.* 2010. [Google Scholar] [CrossRef]

[46] Saranwong, S.; Somsrivicchai, J.; Kawano, S. On-tree evaluation of harvesting quality of mango fruit using a hand-held NIR instrument. *J. Near Infrared Spectrosc.* 2003, 11, 283–293. [Google Scholar] [CrossRef]

[47] Jha, S.N.; Kingsly, A.R.P.; Chopra, S. Non-destructive determination of firmness and yellowness of mango during growth and storage using visual spectroscopy. *Biosyst. Eng.* 2006, 94, 397–402. [Google Scholar] [CrossRef]

[48] Schmilovitch, Z.; Mizrach, A.; Hoffman, A.; Egozi, H.; Fuchs, Y. Determination of mango physiological indices by near-infrared spectrometry. *Postharvest Biol. Technol.* 2000, 19, 245–252. [Google Scholar] [CrossRef]

[49] Mendoza, F.; Dejmek, P.; Aguilera, J.M. Calibrated color measurements of agricultural foods using image analysis. *Postharvest Biol. Technol.* 2006, 41, 285–295. [Google Scholar] [CrossRef]

[50] Jaiswal, P.; Jha, S.N.; Bharadwaj, R. Non-destructive prediction of quality of intact banana using spectroscopy. *Sci. Hortic.* 2012, 135, 14–22. [Google Scholar] [CrossRef]

[51] Adebayo, S.E.; Hashim, N.; Abdan, K.; Hanafi, M.; Mollazade, K. Prediction of quality attributes and ripeness classification of bananas using optical properties. *Sci. Hortic.* 2016, 212, 171–182. [Google Scholar] [CrossRef]

[52] Rajkumar, P.; Wang, N.; Elmasry, G.; Raghavan, G.S.V.; Gariépy, Y. Studies on banana fruit quality and maturity stages using hyperspectral imaging. *J. Food Eng.* 2012, 108, 194–200. [Google Scholar] [CrossRef]

[53] El-Bendary, N.; El Hariri, E.; Hassanien, A.E.; Badr, A. Using machine learning techniques for evaluating tomato ripeness. *Expert Syst. Appl.* 2015, 42, 1892–1905. [Google Scholar] [CrossRef]

[54] Batu, A. Determination of acceptable firmness and colour values of tomatoes. *J. Food Eng.* 2004, 61, 471–475. [Google Scholar] [CrossRef]

[55] Saad, A.; Ibrahim, A.; El-Biale, N. Internal quality assessment of tomato fruits using image color analysis. *Agric. Eng. Int. CIGR J.* 2016, 18, 339–352. [Google Scholar]

[56] Gastélum-Barrios, A.; López-Bórquez, R.; Rico-García, E.; Toledano-Ayala, M.; Soto-Zarazúa, G. Tomato quality evaluation with image processing: A review. *Afr. J. Agric. Res.* 2011, 6, 3333–3339. [Google Scholar] [CrossRef]

[57] Schouten, R.E.; Huijben, T.P.M.; Tijssens, L.M.M.; van Kooten, O. Modelling quality attributes of truss tomatoes: Linking colour and firmness maturity. *Postharvest Biol. Technol.* 2007, 45, 298–306. [Google Scholar] [CrossRef]

[58] Clément, A.; Dorais, M.; Vernon, M. Nondestructive measurement of fresh tomato lycopene content and other physicochemical characteristics using visible NIR spectroscopy. *J. Agric. Food Chem.* 2008, 56, 9813–9818. [Google Scholar] [CrossRef] [PubMed]

[59] Clément, A.; Dorais, M.; Vernon, M. Multivariate approach to the measurement of tomato maturity and gustatory attributes and their rapid assessment by vis-NIR spectroscopy. *J. Agric. Food Chem.* 2008, 56, 1538–1544. [Google Scholar] [CrossRef] [PubMed]

[60] Hoffmann, A.M.; Noga, G.; Hunsche, M. Fluorescence indices for monitoring the ripening of tomatoes in pre- and postharvest phases. *Sci. Hortic.* 2015, 191, 74–81. [Google Scholar] [CrossRef]

[61] Liu, C.; Liu, W.; Chen, W.; Yang, J.; Zheng, L. Feasibility in multispectral imaging for predicting the content of bioactive compounds in intact tomato fruit. *Food Chem.* 2015, 173, 482–488. [Google Scholar] [CrossRef] [PubMed]

[62] Long, R.L.; Walsh, K.B. Limitations to the measurement of intact melon total soluble solids using near infrared spectroscopy. *Aust. J. Agric. Res.* 2006, 57, 403–410. [Google Scholar] [CrossRef]

[63] McGlone, V.A.; Fraser, D.G.; Jordan, R.B.; Künemeyer, R. Internal quality assessment of mandarin fruit by vis/NIR spectroscopy. *J. Near Infrared Spectrosc.* 2003, 11, 323–332. [Google Scholar] [CrossRef]

- [64] Greensill, C.V.; Walsh, K.B. Calibration transfer between miniature photodiode array-based spectrometers in the near infrared assessment of mandarin soluble solids content. *J. Near Infrared Spectrosc.* 2002, 10, 27–36. [Google Scholar] [CrossRef]
- [65] Gómez, A.H.; He, Y.; Pereira, A.G. Non-destructive measurement of acidity, soluble solids and firmness of Satsuma mandarin using Vis/NIR-spectroscopy techniques. *J. Food Eng.* 2006, 77, 313–319. [Google Scholar] [CrossRef]
- [66] Guthrie, J.A.; Reid, D.J.; Walsh, K.B. Assessment of internal quality attributes of mandarin fruit. 2. NIR calibration model robustness. *Aust. J. Agric. Res.* 2005, 56, 417–426. [Google Scholar] [CrossRef]
- [67] Crisosto, C.H.; Crisosto, G.M.; Pitenour, M.A. Testing the reliability of skin color as an indicator of quality for early season “Brooks” (*Prunus avium* L.) cherry. *Postharvest Biol. Technol.* 2002, 24, 147–154. [Google Scholar] [CrossRef]
- [68] Lu, R. Predicting firmness and sugar content of sweet cherries using near-infrared diffuse reflectance spectroscopy. *Trans. Am. Soc. Agric. Eng.* 2001, 44, 1265–1271. [Google Scholar] [CrossRef]
- [69] Carlini, P.; Massantini, R.; Mencarelli, F. Vis-NIR measurement of soluble solids in cherry and apricot by PLS regression and wavelength selection. *J. Agric. Food Chem.* 2000, 48, 5236–5242. [Google Scholar] [CrossRef] [PubMed]
- [70] Sánchez, M.T.; De La Haba, M.J.; Benitez-López, M.; Fernández-Novales, J.; Garrido-Varo, A.; Pérez-Marín, D. Non-destructive characterization and quality control of intact strawberries based on NIR spectral data. *J. Food Eng.* 2012, 110, 102–108. [Google Scholar] [CrossRef]
- [71] Amodio, M.L.; Ceglie, F.; Chaudhry, M.M.A.; Piazzolla, F.; Colelli, G. Potential of NIR spectroscopy for predicting internal quality and discriminating among strawberry fruits from different production systems. *Postharvest Biol. Technol.* 2017, 125, 112–121. [Google Scholar] [CrossRef]
- [72] Liu, C.; Liu, W.; Lu, X.; Ma, F.; Chen, W.; Yang, J.; Zheng, L. Application of multispectral imaging to determine quality attributes and ripeness stage in strawberry fruit. *PLoS ONE* 2014, 9, e87818. [Google Scholar] [CrossRef] [PubMed]
- [73] ElMasry, G.; Wang, N.; ElSayed, A.; Ngadi, M. Hyperspectral imaging for nondestructive determination of some quality attributes for strawberry. *J. Food Eng.* 2007, 81, 98–107. [Google Scholar] [CrossRef]
- [74] Liu, M.; Fu, P.; Cheng, R. Non destructive estimation peach SSC and firmness by multispectral reflectance imaging. *N. Z. J. Agric. Res.* 2007, 50, 601–608. [Google Scholar] [CrossRef]
- [75] Tallada, J.G.; Nagata, M.; Kobayashi, T. Non-destructive estimation of firmness of strawberries (*Fragaria × ananassa* Duch.) using NIR hyperspectral imaging. *Environ. Control Biol.* 2006, 44, 245–255. [Google Scholar] [CrossRef]
- [76] Camps, C.; Christen, D. Non-destructive assessment of apricot fruit quality by portable visible-near infrared spectroscopy. *LWT Food Sci. Technol.* 2009, 42, 1125–1131. [Google Scholar] [CrossRef]
- [77] Slaughter, D.C.; Crisosto, C.H. Nondestructive internal quality assessment of kiwifruit using near-infrared spectroscopy. *Semin. Food Anal.* 1998, 3, 131–140. [Google Scholar]
78. Lee, J.; Kim, S.; Seong, K.; Kim, C.; Um, Y.; Lee, S. Quality prediction of kiwifruit based on near infrared spectroscopy. *Korean J. Hortic. Sci. Technol.* 2012, 30, 709–717. [Google Scholar] [CrossRef]
- [79] McGlone, V.A.; Kawano, S. Firmness, dry-matter and soluble-solids assessment of postharvest kiwifruit by NIR spectroscopy. *Postharvest Biol. Technol.* 1998, 13, 131–141. [Google Scholar] [CrossRef]
- [80] Jannok, P.; Kamitani, Y.; Kawano, S. Development of a common calibration model for determining the Brix value of intact apple, pear and persimmon fruits by near infrared spectroscopy. *J. Near Infrared Spectrosc.* 2014, 22, 367–373. [Google Scholar] [CrossRef]
- [81] Wei, X.; Liu, F.; Qiu, Z.; Shao, Y.; He, Y. Ripeness classification of Astringent persimmon using hyperspectral imaging technique. *Food Bioprocess Technol.* 2014, 7, 1371–1380. [Google Scholar] [CrossRef]
- [82] Omar, A.F. Spectroscopic profiling of soluble solids content and acidity of intact grape, lime, and star fruit. *Sens. Rev.* 2013, 33, 238–245. [Google Scholar] [CrossRef]
- [83] Janik, L.J.; Cozzolino, D.; Dambergs, R.; Cynkar, W.; Gishen, M. The prediction of total anthocyanin concentration in red-grape homogenates using visible-near-infrared spectroscopy and artificial neural networks. *Anal. Chim. Acta* 2007, 594, 107–118. [Google Scholar] [CrossRef] [PubMed]
- [84] Agati, G.; D’Onofrio, C.; Ducci, E.; Cuzzola, A.; Remorini, D.; Tuccio, L.; Lazzini, F.; Mattii, G. Potential of a multiparametric optical sensor for determining in situ the maturity components of red and white *Vitis vinifera* wine grapes. *J. Agric. Food Chem.* 2013, 61, 12211–12218. [Google Scholar] [CrossRef] [PubMed]
- [85] Lenk, S.; Buschmann, C.; Pfündel, E.E. In vivo assessing flavonols in white grape berries (*Vitis vinifera* L. cv. Pinot Blanco) of different degrees of ripeness using chlorophyll fluorescence imaging. *Funct. Plant Biol.* 2007, 34, 1092–1104. [Google Scholar] [CrossRef]
- [86] Baiano, A.; Terracone, C.; Peri, G.; Romaniello, R. Application of hyperspectral imaging for prediction of physico-chemical and sensory characteristics of table grapes. *Comput. Electron. Agric.* 2012, 87, 142–151. [Google Scholar] [CrossRef]
- [87] Abu Bakar, B.H.; Ishak, A.J.; Shamsuddin, R.; Wan Hassan, W.Z. Ripeness level classification for pineapple using RGB and HSI colour maps. *J. Theor. Appl. Inf. Technol.* 2013, 57, 587–593. [Google Scholar]
- [88] Guthrie, J.; Walsh, K. Non-invasive assessment of pineapple and mango fruit quality using near infra-red spectroscopy. *Aust. J. Exp. Agric.* 1997, 37, 253–263. [Google Scholar] [CrossRef]
- [89] Chia, K.S.; Abdul Rahim, H.; Abdul Rahim, R. Prediction of soluble solids content of pineapple via non-invasive low cost visible and shortwave near infrared spectroscopy and artificial neural network. *Biosyst. Eng.* 2012, 113, 158–165. [Google Scholar] [CrossRef]
- [90] Infante, R.; Contador, L.; Rubio, P.; Mesa, K.; Meneses, C. Non-destructive monitoring of flesh softening in the black-skinned Japanese plums “Angelino” and “Autumn beaut” on-tree and postharvest. *Postharvest Biol. Technol.* 2011, 61, 35–40. [Google Scholar] [CrossRef]
- [91] Paz, P.; Sánchez, M.T.; Pérez-Marín, D.; Guerrero, J.E.; Garrido-Varo, A. Nondestructive determination of total soluble solid content and firmness in plums using near-infrared reflectance spectroscopy. *J. Agric. Food Chem.* 2008, 56, 2565–2570. [Google Scholar] [CrossRef] [PubMed]
- [92] Mendoza, F.; Aguilera, J.M. Application of Image Analysis for Classification of Ripening Bananas. *Food Eng. Phys. Prop.* 2004, 69, 478–487. [Google Scholar] [CrossRef]
- [93] Olmo, M.; Nadas, A.; Garcia, J.M. Nondestructive Methods to Evaluate Maturity Level of Oranges. *Sens. Nutr. Qual. Food Nondestruct.* 1998, 65, 365–369. [Google Scholar] [CrossRef]
- <sup>[2]</sup> Hamamatsu Photonics, S9706, [https://www.hamamatsu.com/content/dam/hamamatsu-photonics/sites/documents/99\\_SALES\\_LIBRARY/ssd/s9706\\_kpic1060e.pdf](https://www.hamamatsu.com/content/dam/hamamatsu-photonics/sites/documents/99_SALES_LIBRARY/ssd/s9706_kpic1060e.pdf)
- <sup>[3]</sup> Typical progressive change of reflectance spectra at different ripening stages of tomato | AB. Li, J. Lecourt, and G. Bishop, “Advances in Non-Destructive Early Assessment of Fruit Ripeness towards Defining Optimal Time of Harvest and Yield Prediction—A Review,” *Plants*, vol. 7, no. 1, p. 3, Jan. 2018, doi: <https://doi.org/10.3390/plants7010003>
- <sup>[4]</sup> Hamamatsu Photonics, CCD sensors: <https://www.hamamatsu.com/eu/en/product/optical-sensors/image-sensor/ccd-cmos-nmos-image-sensor/line-sensor/for-industry.html>
- <sup>[5]</sup> Hamamatsu Photonics, InGaAs sensors: <https://www.hamamatsu.com/eu/en/product/optical-sensors/image-sensor/ingaas-image-sensor/ingaas-linear-image-sensor.html>
- <sup>[6]</sup> Hamamatsu Photonics, InGaAs area sensor: <https://www.hamamatsu.com/eu/en/product/optical-sensors/image-sensor/ingaas-image-sensor/ingaas-area-image-sensor.html>
- <sup>[7]</sup> L. Smeesters, W. Meulebroeck, S. Raeymaekers, and H. Thienpont, “Optical detection of aflatoxins in maize using one- and two-photon induced fluorescence spectroscopy,” *Food Control*, vol. 51, pp. 408–416, May 2015, doi: 10.1016/j.foodcont.2014.12.003
- <sup>[8]</sup> Hamamatsu Photonics, SXGA InGaAs Camera: <https://www.hamamatsu.com/eu/en/product/cameras/ingaas-cameras/C16741-40U.html>