

Hamamatsu 3rd FDSS Workshop on 8th June 2016

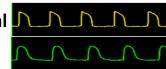


S. BEDUT



For the Institut de Recherche Servier (IdRS), Pôle d'Expertise Biotechnologie, Chimie & Biologie, Croissy-sur-Seine, France



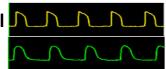


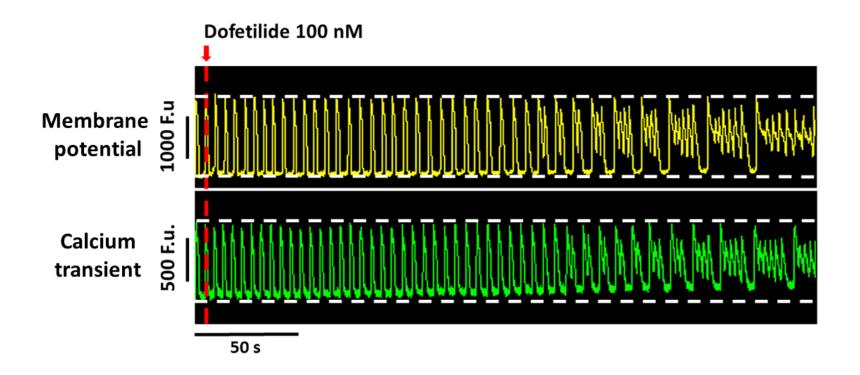
- **1.** Parallel recording of voltage and calcium signals
- 2. FluoVolt dye
- 3. Cellular density (cells/well)
- 4. FluoVolt concentration
- 5. FluoVolt Toxicity
- 6. Washed-out calcium dyes







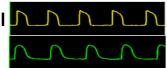




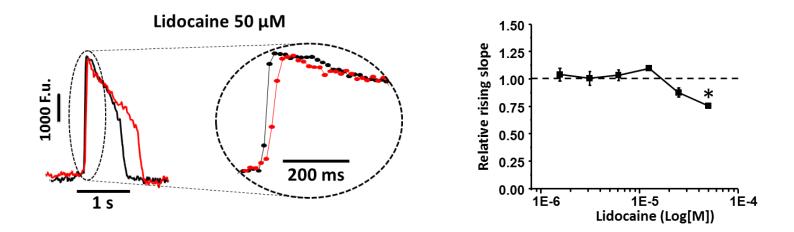












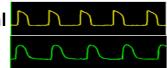




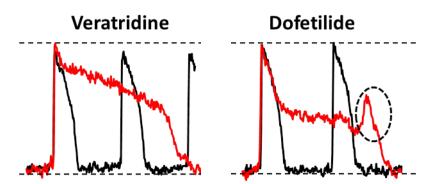


S. BEDUT VWR

Experimental parameters for an efficient recording of membrane potential and calcium activity from hIPS-derived cardiomyocytes in 96 wells

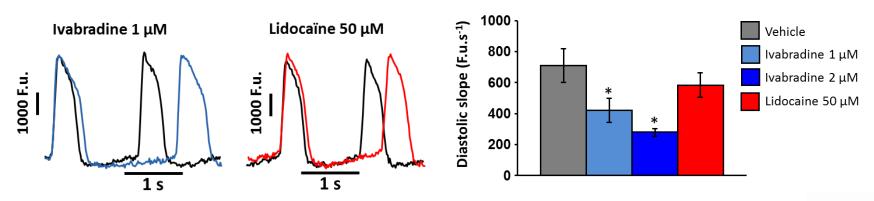


1. Paralell recording of voltage and calcium signals



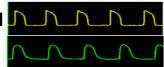
Phase 1 and 3 alteration

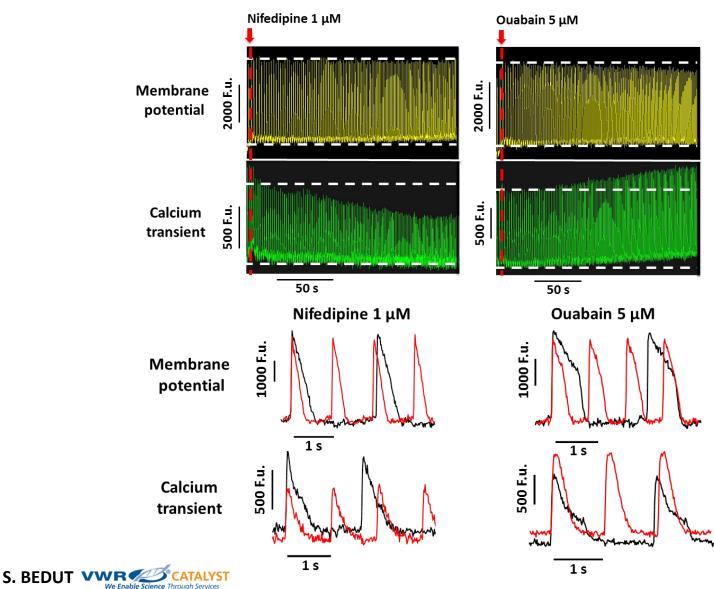
Phase 4 alteration





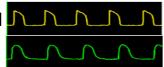


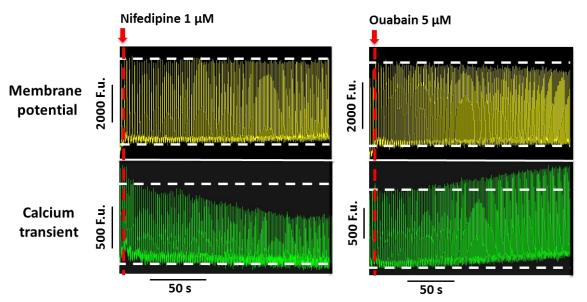


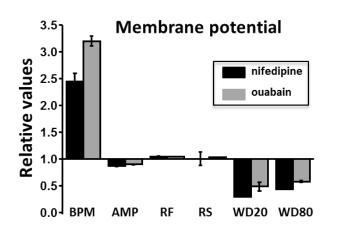








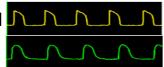


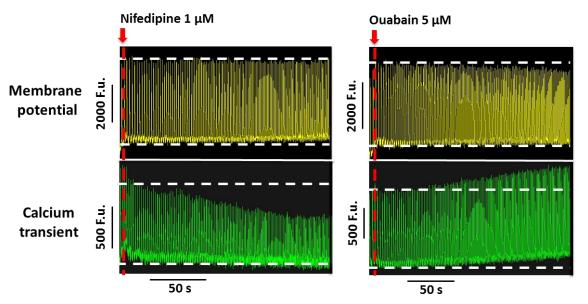


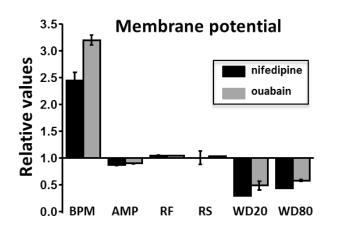


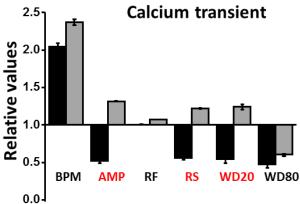








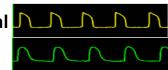






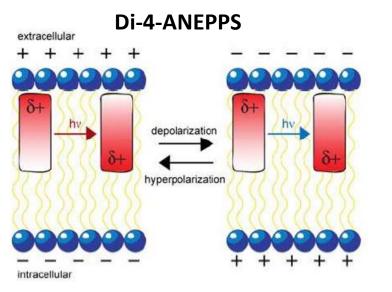






2. FluoVolt Dye

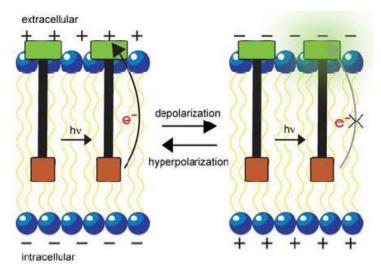
Electrochromic



Shift of the excitation/emission spectra

 Δ F/F \approx 6-10 % per 100 mV Rapid phototoxicity

Photo-induced electron transfer



Modulation of the electron transfer rate

∆F/F ≈ 20-25 % per 100 mV ?

From Miller et al., PNAS 2012, 109 (6) : 2114-2119

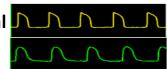






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Experimental parameters for an efficient recording of membrane potential normalized and calcium activity from hIPS-derived cardiomyocytes in 96 wells

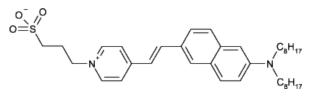


2. FluoVolt Dye

Electrochromic

Di-4-ANEPPS

4-[2-[6-(Dioctylamino)-2-naphthalenyl]ethenyl]-1-(3-sulfopropyl)-pyridinium



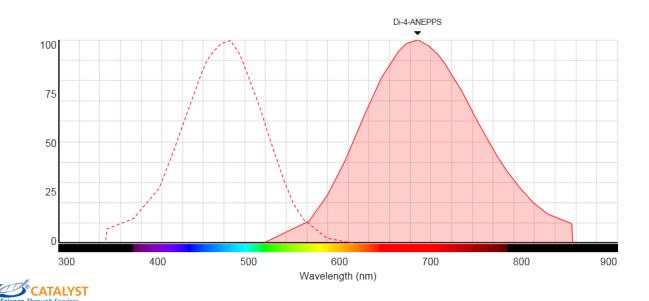
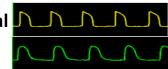




Photo-induced electron transfer

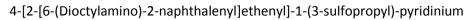


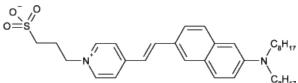


2. FluoVolt Dye

Electrochromic

Di-4-ANEPPS





100

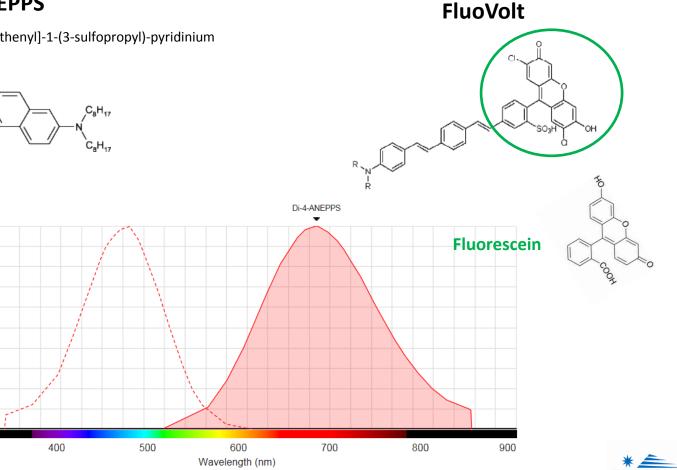
75

50

25

300

Photo-induced electron transfer







o⁻ // o=s

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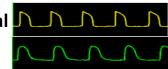


Photo-induced electron transfer

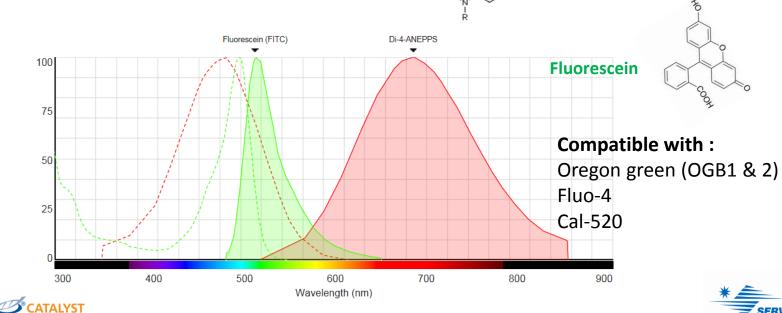
2. FluoVolt Dye

Electrochromic

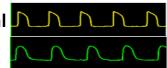
Di-4-ANEPPS



FluoVolt







2. FluoVolt Dye

96 wells

Experimental media: HBSS + 20 mM Hepes, pH 7.4 with NaOH (serum-free, phenol red-free)

Temp: 37°C

Loading time: 15 mn (50 µl, 37°C)

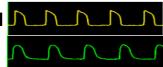
Rinced twice (2 x 100 µl)

Sampling interval: 16 ms

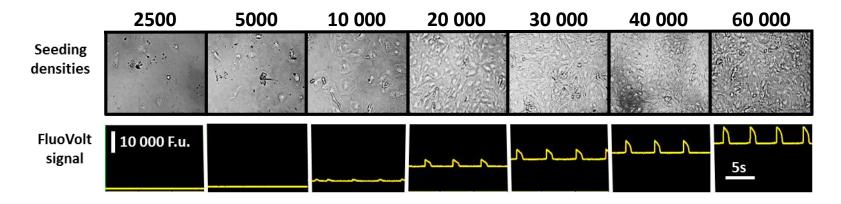


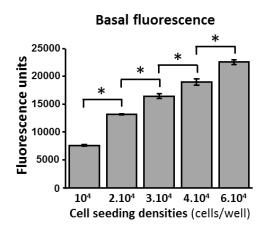






3. Cellular density (cells/well)

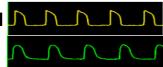




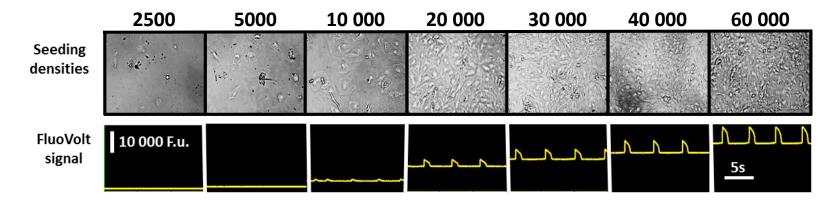




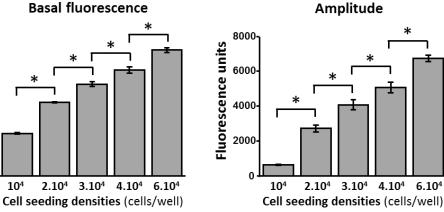




3. Cellular density (cells/well)









25000

20000

15000

10000

5000

n

10⁴

Fluorescence units

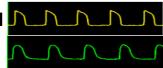




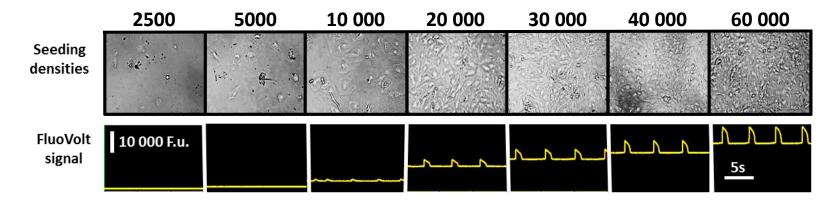
Fluorescence units

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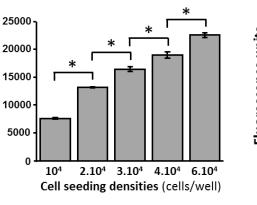


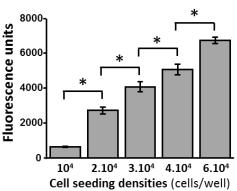
3. Cellular density (cells/well)





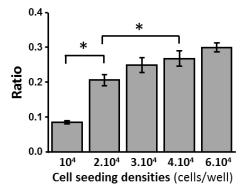
FALYST





Amplitude





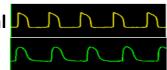






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Experimental parameters for an efficient recording of membrane potential and calcium activity from hIPS-derived cardiomyocytes in 96 wells

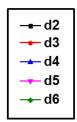


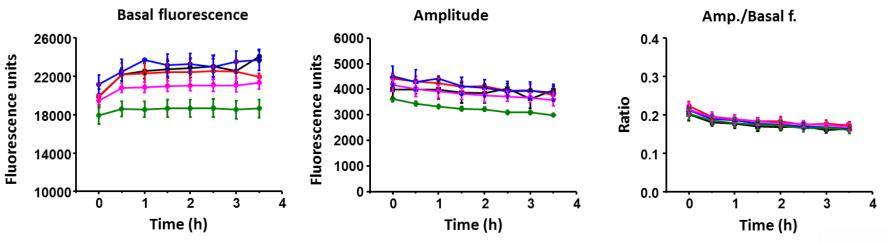
4. FluoVolt concentration



In 10, 20, 30, 40, 50 and 60 ml HBSS (d1 to d6) d1, d2, d3, d4, d5 and d6

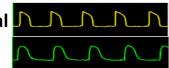
50 μ l + 500 μ l Powerload







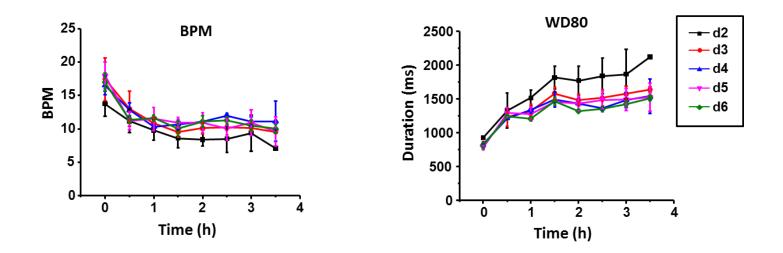




5. FluoVolt toxicity



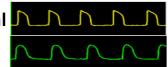
In 10, 20, 30, 40, 50 and 60 ml (d1 to d6) d1, d2, d3, d4, d5 and d6











6. Washed-out calcium dyes

For an easy to operate assay, the same protocol as FluoVolt is expected

96 wells

Experimental media: HBSS + 20 mM Hepes, pH 7.4 with NaOH

Temp: 37°C

Loading time: 15 mn (50 µl, 37°C) Oregongreen

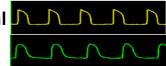
Rinced twice (2 x 100 μl) Dye extrusion

Sampling interval: 16 ms

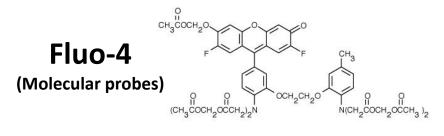


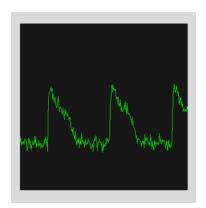




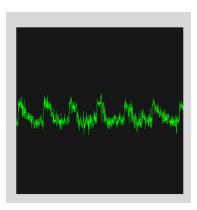


6. Washed-out calcium dyes





Start

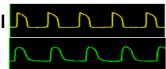


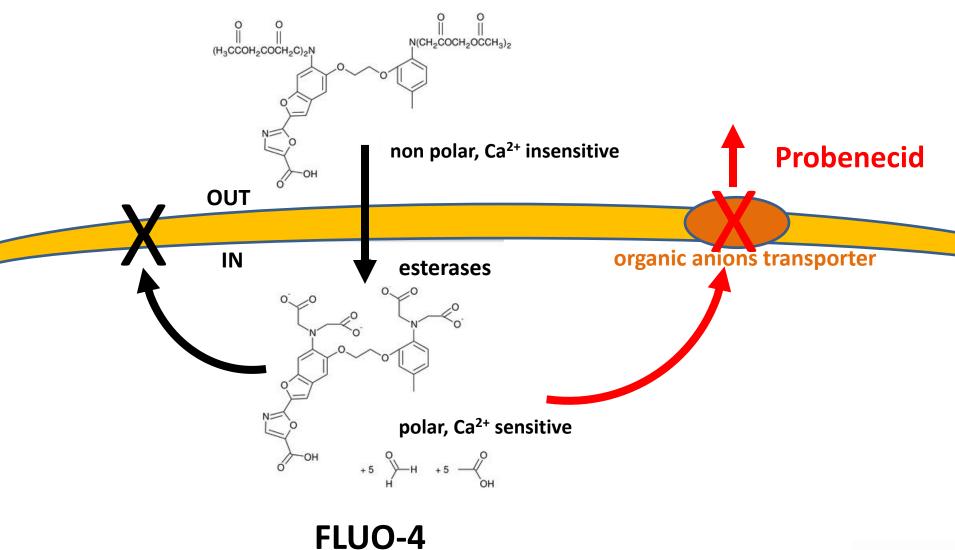
30 mn











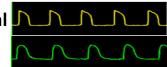


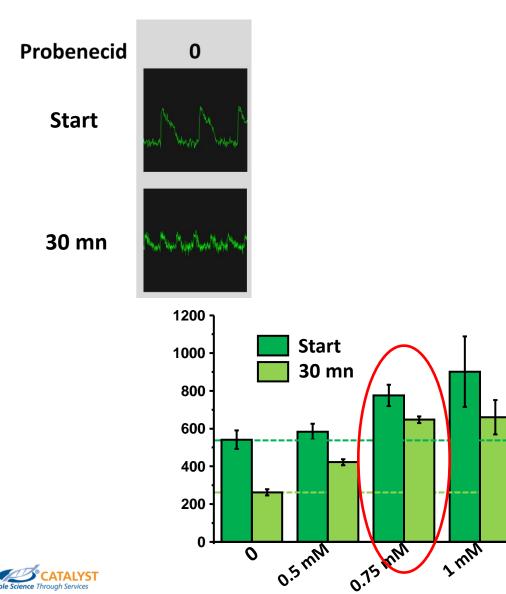




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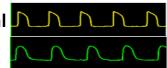
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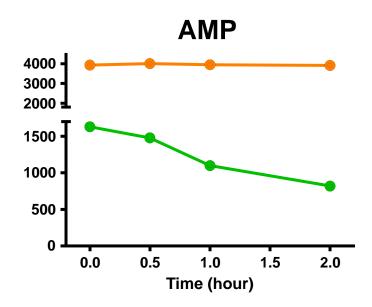


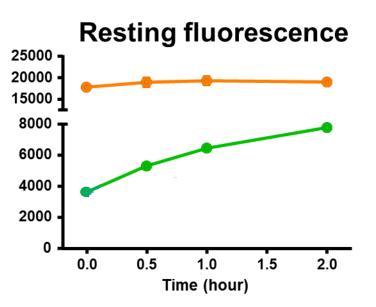








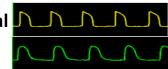


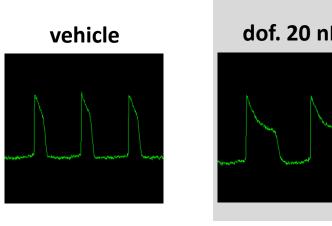


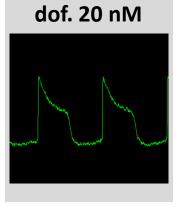


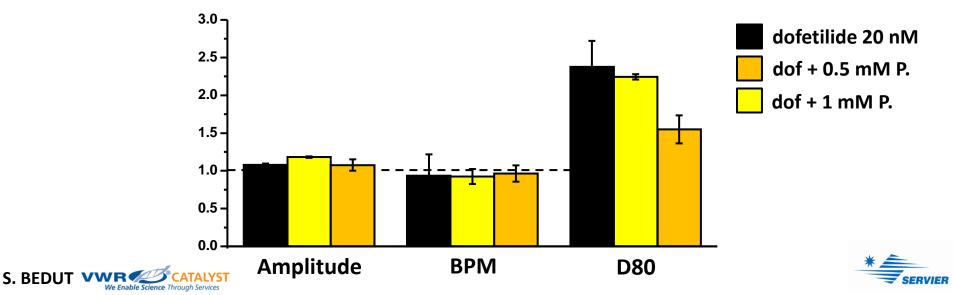




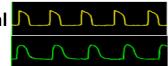






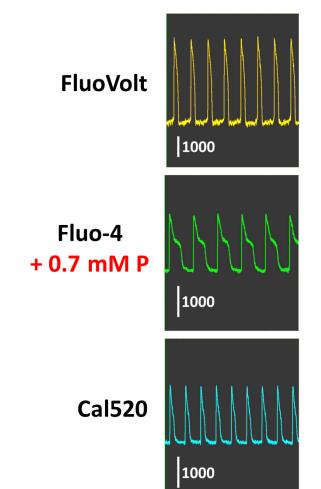






6. Washed-out calcium dyes

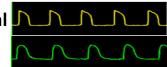
Cal-520 (AAT bioquest)



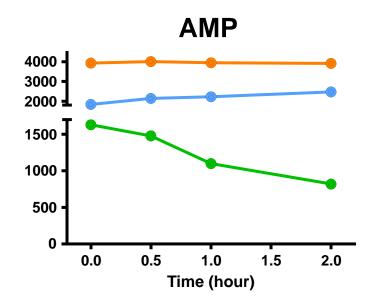








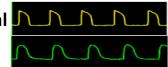


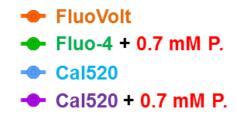


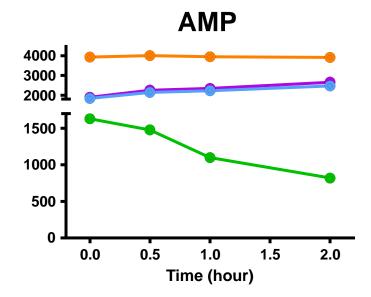


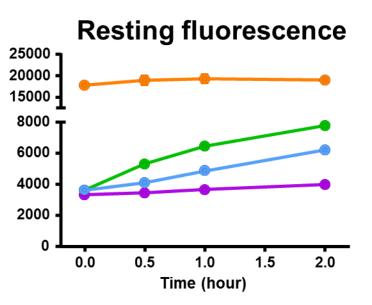








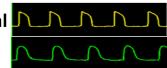












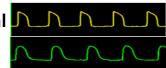
7. Conclusions

No serum, no phenol-red (HBSS + Hepes) No probenecid High cellular density 37°C 16 ms sample interval FluoVolt Cal-520 15 mn loading time 2 wash-outs About 3 hours of recording capacity

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H. MYSKOWSKI



J.P. STEPHAN F. COGE V. LAMAMY N. VILLAIN

Thank you for your attention



